## Therapeutic Potential of Iron Chelating Drugs

Chaim Hershko, Abraham M Konijn and Gabriela Link

ped dispersion of the second o

Department of Medicine, Shaare Zedek Medical Center and Department of Human Nutrition and Metabolism, Hebrew University Hadassah Medical School, Jerusalem, Israel P O Box 3235

Correspondence: C Hershko, Department of Medicine, Shaare Zedek Medical Center Jerusalem, Israel P O Box 3235 E-mail hershko@szmc.org.il



CONFERENCIA

inequipment in the literal

neritta Dos taves

15 15 15 19 11

HEMATOLOGIA, Vol. 3 N° 2: 67-71 Mayo - Octubre, 1999

The primary abnormality in thalassemia major is a wasteful, ineffective erythropoiesis resulting in a 10 to 15-fold expansion of the erythroid bone marrow (1) and a drastic increase in hemoglobin catabolism. Iron accumulation is the consequence of blood transfusions as well as of increased iron absorption caused by erythropoietic activity. The combination of iron overload and increased outpouring of catabolic iron from the reticuloendothelial system overwhelms the iron carrying capacity of transferrin, resulting in the emergence of toxic non-transferrin bound plasma iron (NTPI, 2-4).

Recent studies by Porter et al (5) have shown that plasma NTPI is removed by intravenous deferoxamine (DF) therapy in a biphasic manner and that upon cessation of DF infusion it reappears rapidly, lending support to the continuous, rather than intermittent, use of DF in high risk patients. The rate of low molecular weight iron uptake by cultured rat heart cells is over 300-times that of transferrin iron (6). Moreover, unlike transferrin-iron uptake which is inhibited at high tissue iron concentrations by down-regulation of transferrin receptor production, non-transferrin iron uptake is increased by high tissue iron (7). Such uptake results in increased myocardial lipid peroxidation and abnormal contractility, and these effects are reversed by in vitro treatment with DF (8). Recognition of NTPI as a potentially toxic component of plasma iron in thalassemic siderosis has important practical implications for designing better strategies for the effective administration of DF and other iron chelating drugs.

In thalassemic patients who are not receiving iron chelation therapy, the accumulation of iron will progress relentlessly and when about 20 grams of

iron have been acquired, severe clinical manifestations of iron toxicity may be anticipated (9). The most important complications of transfusional siderosis are cardiac, hepatic and endocrine disease. Pathologic findings in the heart include dilated, thickened ventricular walls with particularly heavy iron deposits in the ventricles, epicardium and papillary muscles. These cellular deposits induce increased membrane lipid peroxidation in the sarcolemma resulting in impaired Na,K,ATPase activity (10), increased lysosomal fragility (11) and, in particular, impaired mitochondrial inner-membrane respiratory chain activity (12). It is possible to demonstrate early myocardial dysfunction in asymptomatic patients using MUGA scan (13) or dobutamine stress echocardiography (14). Advanced cardiac siderosis results in heart failure and life-threatening arrhythmias. Myocardial siderosis is the single most important cause of mortality in inadequately treated thalassemic patients.

Cirrhosis is a common complication of thalassemia and, similar to cardiac problems, its incidence is age-related. However, the coexistence of chronic hepatitis B or C with an incidence ranging from 9 to 70% of thalassemic patients in various geographic areas (9) underlines the complexity of this problem. Iron overload per se is responsible for the development of cirrhosis in many cases. It has been proposed, that patients free of hepatitic virus infection, with liver iron concentrations below 15 mg/gm dry weight may have a prolonged survival free of the clinical complications of iron overload (15). Endocrine problems caused by direct accumulation of iron in endocrine glands or indirectly through the hypothalamic-pituitary axis are common. Stunted growth, delayed puberty, hypothyroidism, hypoparathyroidism and diabetes mellitus are all well established complications of transfusional siderosis (16). Because diabetes and hypothyroidism appear when most endocrine cells are destroyed and replaced by fibrosis, these complications are rarely reversible.

## RESULTS OF DEFEROXAMINE THERAPY

Although the impact of DF therapy on the survival and well-being of thalassemic patients has never been proven by prospective, randomized clinical studies, the beneficial effects of long-term deferoxamine treatment are clearly demonstrated by comparison of treatment outcome with historical controls. Experience with long-term DF therapy in thalassemic patients has been summarized in several extensive recent reviews (9,17,18)

Iron chelating treatment should be started when serum ferritin levels reach about 1000 mg/L which usually occurs after the first 10 or 20 transfusions (17). DF is infused via a thin s.c. needle inserted to the arm or abdomen nightly, connected to a portable pump over 8-12 h, 5 to 7 times per week at a daily dose of 20 to 60 mg/kg. A urinary iron excretion of 0.5 mg/kg/d is usually sufficient to ensure negative iron balance. A new delivery system for continuous DF infusion has been introduced by Baxter allowing continuous 48 h s.c. or continuous 24 h i.v. delivery for 7 days each week (19). This technology allows effective removal of toxic free iron (NTPI) from the plasma, a significant decrease in serum ferritins within 4 weeks, and improves patient compliance compared to conventional s.c. DF pumps.

Recently, a depot preparation of DF has been developed by Novartis which, by delivering a smaller dose over a longer period of time, makes it more efficient and reduces the proportion of non-chelated wasted DF which is up to 90% of the drug delivered by current methods (20). Preliminary studies have shown that the duration of action of the depot formulation is over 30 hours, and cumulative iron excretion is more than 3 times that of standard s.c. bolus DF injections. If local torelability of the new depot preparation in current clinical studies proves to be acceptable, this new technology may permit once-daily or alternate day s.c. injections, obviating the need for portable pumps and improving patient compliance.

Response to treatment may be assessed by serum ferritin measurements, liver biopsies, computed tomography, or magnetic susceptibility (SQUID) (21). Serum ferritins are disproportionately low in patients with coexistent ascorbate deficiency (22) and high in

active liver disease or inflammation. Nevertheless, serum ferritin is the most accessible and inexpensive tool for the long-term monitoring of chelating efficiency and protection from cardiac complications may be achieved when ferritin levels are kept below 2,500 mg/L (23).

The impact of deferoxamine treatment on life expectancy is convincingly demonstrated by comparison of survival in well chelated versus poorly chelated patients. In a major study of 1127 thalassemic patients at 7 Italian teaching hospitals, it was shown that 70% of patients born before 1970 and hence prior to the modern era of chelation survived to the age 20 y. compared to 88% in patients born after 1970 and therefore receiving effective chelation from an early age (9). Most of the improvement in survival was attributed to decreased cardiac mortality. This cohort-of-birth related improvement in survival is reflected in a mirror-like inverse decrease in cardiac mortality, supporting the assumption that prevention of cardiac mortality is the most important beneficial effect of DF therapy. Improved survival in well chelated thalassemic patients has been reported in several other major studies from the U.K. (24) and North America (15,23,25).

The strongest direct evidence supporting the beneficial effect of DF on hemosiderotic heart disease is the reversal of established myocardiopathy in some far-advanced cases. Earlier experience in hereditary hemochromatosis has shown that the myocardiopathy of iron overload is potentially curable by effective iron mobilization through phlebotomy. However, in transfusional hemosiderosis, the course of established myocardial disease was uniformly fatal and, until recently, believed to be non-responsive to iron chelating therapy. Several reports indicate that such patients may still be responsive to aggressive chelating treatment. Marcus et al (26) described first the reversal of established symptomatic myocardial disease in 3 of 5 patients by continuous high-dose (85-200 mg/kg/d) i.v. DF therapy at the cost of severe reversible retinal toxicity. Reversal of symptomatic myocardiopathy has been reported by others, without significant drug toxicity (27,28). Continuous 24hour ambulatory intravenous infusion of DF through central venous ports, using standard portable infusion pumps or the new Baxter delivery system is a very effective method for the rapid reversal of established hemosiderotic heart disease. In addition, it is an excellent tool for improving patient compliance allowing uninterrupted delivery of 6 to 12 grams DF per day and the effective depletion of very large iron stores.

## CONDITIONS UNRELATED TO IRON OVERLOAD

Because iron plays a central role in many important biological reactions such as the formation of toxic oxygen species, mitochondrial inner membrane respiratory complex activity, and the activity of ribonucleotide reductase, a rate limiting enzyme in cell replication, iron chelators have a potential role as therapeutic agents in conditions wherein interference with the above functions may modify the pathogenetic process. Because of limitations of space, I shall limit this part of my review to the effect of iron chelators on intracellular parasites.

The antimalarial effect of iron chelators: A number of experimental and clinical studies indicate that iron deficiency may have an important inhibitory effect on the progression of malarial infection and, conversely, that iron repletion may result in the exacerbation of malaria. However, this hypothesis is not universally accepted, as other studies have been unable to show an adverse effect of iron administration on human malaria (29), and severe iron deficiency may interfere with the normal immune response thus aggravating, rather than inhibiting infection.

In view of the possible beneficial effects of iron depletion, DF has been studied as a potential antimalarial agent. DF inhibits the growth of P. falciparum cultures at concentrations above 20 mM (30). In vivo studies in rats infected with P berghei, mice with P vinckei and monkeys with P falciparum have shown that DF is able to suppress malaria if a continuous supply of the chelator is assured by frequent (8 hourly) subcutaneous injections (31,32), or by osmotic pumps.

Encouraged by these studies in experimental animals, several investigators have tested the antimalarial effect of DF in humans. Traore et al (33) have studied the effect of DF 0.5 g i.m. given twice daily for 3 days on the rate of clearance of parasitemia in patients with P. falciparum malaria who were also receiving chloroquine. Although parasitemia appeared to decrease more rapidly in the 6 patients receiving DF and chloroquine than in the 3 controls treated by chloroquine only, the small number of patients, and the inclusion of chloroquineresistant cases with resurgent malaria limit the value of this preliminary report. In another clinical study by Bunnag et al (34) 14 patients with symptomatic P. vivax and 14 with uncomplicated P. falciparum malaria received continuous i.v. DF 100 mg/kg for 72 hours. No other antimalarial treatment was given. In both groups DF reduced the parasitemia to zero within 57 to 106 hours. There was significant drug toxicity with transient visual blurring in 9 patients. Recrudescence was observed within the subsequent 3 weeks in all but 2 patients. A major weakness of this study was the absence of a control group.

Two controlled studies of DF in human malaria have been conducted by Gordeuk et al. In the first of these, the effect of DF therapy in partially immune adults with asymptomatic P. falciparum parasitemia has been tested (35,36). A detailed description of these important clinical studies can be found in a subsequent chapter of this volume.

Collectively, these studies leave no doubt as to the ability of DF to hasten recovery from malaria, presumably by inhibiting parasite growth in a similar fashion to its effect in experimental in vitro and in vivo systems. In cerebral malaria, an additional beneficial effect could be inhibition of oxidative brain damage by preventing the formation of toxic free radicals through the iron-driven Fenton reaction. However, as emphasized in several recent editorials (37), additional large-scale carefully controlled studies are needed, with particular emphasis on mortality and neurological sequelae, before DF could be recommended for the treatment of cerebral malaria.

In order to explore the role of lipophilicity in antimalarial activity, we have examined the antimalarial effects of 3-hydroxypyrid-4-ones (38), a family of bidentate orally effective iron chelators. All 3-hydroxypyrid-4-ones have an identical stability constant for iron(III), but they may be made more, or less lipophilic by increasing or reducing the length of the R, substituent on the ring nitrogen. Of the hydroxypyridin-4-ones investigated in our studies), those with the highest lipid solubility proved to be the most efficient antimalarial compounds. Subsequent studies by Shanzer et al (39) employing a series of synthetic iron chelators have confirmed our conclusions that the antimalarial effect of iron chelators is determined by their lipophilicity as well as their affinity to iron.

Other studies have shown that DF is able to inhibit the proliferation in vitro and in vivo of Leishmania donovani (40), Trypanosoma cruzi (41), Pneumocystis carinii (42), and Legionella pneumophila (43). These intriguing observations on the antimicrobial effects of DF and other iron chelators lend new meaning to the term «Nutritional Immunity» (44) and open new channels for exploring the possibility of controlling infection by means of selective intracellular iron deprivation. Experimental models for studying the effect of iron chelators on other intracellular pathogens such as Toxoplasma gondii, Chlamidia psittaci, or Mycobacterium tuberculosis should be established. Packaging the chelator in liposomes or red cell ghosts, or manipulating their lipid solubility to improve their delivery to appropriate target organs such as the

macrophage system may greatly improve their efficiency. In view of the short half-life and poor oral effectiveness of DF, it is unlikely that this drug will be suitable for clinical use as a practical antimicrobial agent. However, with the introduction of simple, orally effective new chelators, it is reasonable to expect that future research may lead to the identification of iron chelators with considerable usefulness in the control of infectious disease.

## REFERENCES

- Hershko C, Rachmilewitz EA. Mechanism of desferrioxamineinduced iron excretion in thalassaemia. Brit J Haematol, 1979, 42, 125-132.
- Hershko C, Graham G, Bates GW, Rachmilewitz EA. Nonspecific serum iron in thalassaemia: an abnormal serum iron fraction of potential toxicity. Brit J Haematol, 1978, 40, 255-263.
- Gutteridge JMC, Rowley DA, Griffiths E, Halliwell B. Lowmolecular-weight iron complexes and oxygen radical reactions in idiopathic haemochromatosis. Clin Sci 1985, 68, 463-467.
- Al-Refaie FN, Wickens DG, Wonke B, Kontoghiorghes GJ, Hoffbrand AV. Serum non-transferrin-bound iron in betathalassaemia major patients treated with desferrioxamine and L1. Brit J Haematol, 1992, 82, 431-436.
- Porter JB, Abeysinghe RD, Marshall L, Hider RC, Singh S. Kinetics of removal and reappearance of non-transferrin-bound plasma iron with deferoxamine therapy. Blood, 1996, 88, 705-713.
- Link G, Pinson A, Hershko C. Heart cells in culture: a model of myocardial iron overload and chelation. J Lab Clin Med, 1985, 106, 147-153.
- Randell EW, Parkes JG, Olivieri NF, Templeton DM. Uptake of non-transferrin-bound iron by both reductive and nonreductive processes is. modulated by intracellular iron. J Biol Chem, 1994, 269, 16046-16053.
- Link G, Athias P, Grynberg A, Pinson A, Hershko C. Effect of iron loading on transmembrane potential, contraction and automaticity of rat ventricular muscle cells in culture. J Lab Clin Med, 1989, 113, 103-111.
- Gabutti V, Borgna-Pignatti C. Clinical manifestations and therapy of transfusional haemosiderosis. Clin Haematol, 1994, 7, 919-940.
- Link G, Pinson A, Hershko C. The ability of orally effective iron chelators dimethyl- and diethyl-hydroxypyrid-4-one and of deferoxamine to restore sarcolemmal thiolic enzyme activity in iron-loaded heart cells. Blood, 1994, 83, 2692-2697.
- Link G, Pinson A, Hershko C. Iron loading of cultured cardiac myocytes modifies sarcolemmal structure and increases lysosomal fragility. J Lab Clin Med, 1993, 121, 127-134.
- Link G, Tirosh R, Pinson A, Hershko C. Role of iron in the potentiation of anthracycline toxicity: Identification of heart cell mitochondria as the site of iron-anthracycline interaction. J Lab Clin Med, 1996, 127, 272-278.
- Aldouri MA, Wonke B, Hoffbrand AV, Flynn DM, Ward SE, Agnew JE et al High incidence of cardiomyopathy in betathalassaemia patients receiving regular transfusion and iron chelation: reversal by intensified chelation. Acta Haematol 1990, 83, 113-117.
- Mariotti E, Agostini A, Angelucci E. Cardiac study by dobutamine stress echocardiography in thalassemic patients. Bone Marrow Transpl, 1993, 12, (suppl 1): 14-15.
- 15. Brittenham GM, Griffith PM, Nienhuis AW. Efficacy of

- deferoxamine in preventing complications of iron overload in patients with thalassemia major. New Engl J Med, 1994, 331, 567-569.
- De Sanctis V, Pintor C, Aliquo MC. Prevalence of endocrine complications in patients with b-thalassemia major: an Italian multicenter study. In: C. Pintor, E.E. Muller, S. Loche & M.I. New (eds). Advances in Pediatric Endocrinology 1992, pp 127-133, Milano, Springer-Verlag, Berlin.
- Gabutti V, Piga A. Results of long-term iron-chelating therapy. Acta Haematol, 1996, 95, 26-36.
- Olivieri NF, Brittenham GM. Iron chelating therapy and the treatment of thalassemia. Blood, 1997, 89, 739-761.
- Araujo A, Kosaryan M, MacDowell A, Wickens D, Puri S, Wonke B et al A novel delivery system for continuous desferrioxamine infusion in transfusional iron overkload. Brit J Haematol, 1996, 93, 835-837.
- Alberti D, Piga A, Porter JB, Hassan I. Desferrioxamine depot (CGH 749B): a new formulation allowing the prospect of a daily s.c. injection. (Abstract) 8th International Conference, Oral Chelation in the Treatment of Thalassaemia and Other Disease. Corfu. 1997, p.39.
- Brittenham GM, Farrell DE, Harris DE, Feldman ES, Danish EH, Muir DA et al. Magnetic susceptibility measurement of human iron stores. New Engl J Med 1982, 307, 1671-1675.
- Chapman RWG, Hussain MAM, Gorman A, Laulicht M, Politis, Flynn DM, et al. Effect of ascorbic acid deficiency on serum ferritin concentration in patients with beta thalassemia. J Clin Pathol, 1982, 35, 481-486.
- Olivieri NF, Nathan DG, MacMillan JH, Wayne AS, Liu PP, McGee A, et al. Survival in medically treated patients with homozygous b-thalassemia. New Engl J Med 1994, 331, 574-578.
- Hoffbrand AV, Wonke B. Results of long-term subcutaneous desferrioxamine therapy. Bailliere's Clin Haematol, 1989, 2, 345-359.
- Giardina PJ, Grady RW, Ehlers KH, Burstein S, Graziano JH, Markenson AL, et al. Current therapy of Cooley's anemia. A decade of experience with subcutaneous desferrioxamine, In: A. Bank, Ed. Sixth Cooley's Anemia Symposium, Ann N Y Acad Sci 1990, 612, 275-285.
- Marcus RE, Davies SC, Bantock HM, Underwood SR, Walton S, Huehns ER. Desferrioxamine to improve cardiac function in iron-overloaded patients with thalassaemia major. Lancet, 1984, 1, 392.
- Hyman CB, Agness CL, Rodriguez-Funes R, Zednikova M. Combined subcutaneous and high-dose intravenous deferoxamine therapy of thalassemia, Ann N Y Acad Sci 1985, 445, 293-303.
- Cohen AR, Martin M, Schwartz E. Current treatment of Cooley's anemia. Intravenous chelation therapy. In: A. Bank, Ed. Sixth Cooley's Anemia Symposium, Ann N Y Acad Sci, 1990, 612, 286-292.
- Harvey P, Heywood P, Nesheim MC, Habicht JP, Alpers M: Iron repletion and malaria. Fed Proc 1987, 46, 1161. (abstr).
- Raventos-Suarez C, Pollack S, Nagel RL: Plasmodium falciparum:inhibition of in vitro growth by desferrioxamine. Amer J Trop Med Hyg 1982, 31, 919-922.
- Hershko C, Peto TEA: Deferoxamine -inhibition of malaria is independent of host iron status. J Exper Med 1988, 168, 375-387.
- Fritch G, Treumer J, Spira DT, Jung A: Plasmodium vinckei: Suppression of mouse infections with desferrioxamine B. Exper Path 1985, 60, 171-174.
- Traore O, Carnevale P, Kaptue-Noche L et al: Preliminary report on the use of desferrioxamine in the treatment of Plasmodium falciparum malaria. Amer J Hemat 1991, 37, 206-208.
- Bunnag D, Poltera AA, Viravan C et al: Plasmodicidal effect of

- desferrioxamine B in human vivax and falciparum malaria from Thailand. Acta Trop Basel 1992, 52, 59-67.
- Gordeuk VR, Thuma PE, Brittenham GM, et al: Iron chelation with desferrioxamine B in adults with asymptomatic Plasmodium falciparum parasitemia. Blood 1992, 79, 308-312.
- Gordeuk VR, Thuma P, Brittenham GM et al: Effect of iron chelation therapy on recovery from deep coma in children with cerebral malaria. New Engl J Med 1992, 327, 1473-1477.
- Wyler DJ: Bark, weeds, and iron chelators Drugs for malaria.
   New Engl J Med 1992, 327, 1519-1521.
- Hershko C, Theanacho EN, Spira DT et al: The effect of N-alkyl modification on the antimalarial activity of 3-hydroxypyrid-4one oral iron chelators. Blood 1991, 77, 637-643.
- Shanzer A, Libman J, Lytton S, Glickstein H, Cabantchik ZI: Reversed siderophores act as antimalarial agents. Proc Natl Acad Sci 1991, 88, 6585-6589.

- Segovia M, Navarro A, Artero JM: The effect of liposomeentrapped desferrioxamine on Leishmania donovani in vitro. Ann Trop Med Parasitol 1989, 83, 357-360.
- Lalonde RG, Holbein BE: Role of iron in Trypanosoma cruzi infection in mice. J Clin Invest 1984, 73, 470-476.
- Clarkson AB, Saric S, Grady RW: Deferoxamine and effornitine (DF-a-difluoromethylornithine) in a rat model of Pneumocystis carinii pneumonia. Antimicr Agent Chemother 1990, 34, 1833-1835.
- Byrd TF, Horwitz MA: Interferon gamma-activated human monocytes downregulate transferrin receptors and inhibit the intracellular multiplication of Legionella pneumophila by limiting the availability of iron. J Clin Invest 1989, 83, 1457-1465.
- Kochan I: The role of iron in bacterial infections with special considerations of host-tubercle bacillus interaction. Curr Topics Microbiol Immunol 1973, 60, 1-30.