Thalassemia – State-of-the-Art, 2007

Eliezer A. Rachmilewitz



CONFERENCIA

the Edith Wolfson Medical Centrer, Holon, Israel

HEMATOLOGIA, Vol. 11 N° 2: 51-55 Mayo - Octubre, 2007

ABSTRACT

Thalassemia is a hereditary anemia resulting from defects in hemoglobin production.

 β or α -Thalassemia, which is caused by a decrease in the production of α or β -globin chains, affects multiple organs and is associated with considerable morbidity and mortality. Accordingly, lifelong care is required, and financial expenditures for proper treatment are substantial.

Thalassemia is among the most common genetic disorders worldwide; 4.83 percent

of the world's population carry globin variants, including 1.67 percent of the population who are heterozygous for α -thalassemia and β -thalassemia.

 β -Thalassemia is caused by any of more than 200 point mutations and, rarely, by deletions, while in α -thalassemia 50 deletions and about 100 point mutations have been identified. Thalassemia is clinically heterogeneous because various genetic lesions variably impair globin-chain synthesis. However, genotypic variability at known loci is often insufficient to explain the disparate phenotypes of individual patients with the same genotype. Disparity between genotypes and phenotypes is particularly marked in β thalassemia intermedia where patients are not regularly transfused like those with β -thalassemia major, and hemoglobin E thalassemia.

Hemolysis and ineffective erythropoiesis together cause the anemia that occurs in thalassemia. The relative contributions of these two pathologic processes differ in various forms of thalassemia and is mainly related to the quantity of the remaining globin chains. In β -thalassemia, the unstable α globin chains precipitates much faster than excess β globin chains in α

thalassemia which results in premature denaturation of the affected red blood cells, with al the deleterious consequences.

Regular transfusion therapy to maintain hemoglobin levels of at least 9 to 10 g per deciliter allows for improved growth and development and also reduces hepatosplenomegaly due to extramedullary hematopoiesis as well as bone deformities.

Impairment of growth and endocrinopathies, particularly hypogonadism and "bronze diabetes", are common features of thalassemia particularly in older patients or in those where iron chelation therapy is insufficient. Considerable morbidity in older patients results from bone disease due to osteopenia and osteoporosis, which is often accompanied by disabling pain and fractures. The pathogenesis is complex and multifactorial. Bone marrow expansion due to ineffective erythropoiesis, endocrine dysfunction, and complications of treatment all contribute to the condition.

Iron overload causes most of the mortality and morbidity associated with thalassemia. Iron deposition occurs in visceral organs (mainly in the heart, liver, and endocrine glands), causing tissue damage and ultimately organ dysfunction and failure. Cardiac events due to iron overload are still the primary cause of death. Both transfusional iron overload and excess gastro-intestinal absorption are contributory.

Accurate, preferably noninvasive measurement of iron stores is crucial for the evaluation and management of chelation therapy. Serum ferritin is most commonly measured as an indicator of iron stores. Ferritin levels below 2500 ng per milliliter are associated with improved survival. The application of T2 gradientecho sequencing is more sensitive to hemosiderin de-

position and appears to be useful for the measurement mainly of myocardial and liver iron in thalassemia. Elevated tissue iron stores are only one component of the damaging effects of iron overload. A highly toxic form of iron, non-transferrin-bound iron, is formed when the iron-binding capacity of transferrin has been exceeded. Non-transferrin bound iron is highly toxic because it can catalyze the formation of reactive oxygen species through the Fenton reaction. Evidence for oxidative stress has been found in all 3 blood cell components and in major organs.

Iron-chelation therapy is largely responsible for doubling the life expectancy of patients with thalassemia major. Deferoxamine continues to be the most common iron-chelating agent in use, despite the less convenient parenteral administration. Deferiprone, an oral iron chelator, has a number of advantages over deferoxamine. It can penetrate the cell membrane and chelate toxic intracellular iron species and may be more effective than deferoxiamine in the removal of myocardial iron. An encouraging new approach to chelation therapy is the sequential combined administration of deferiprone and deferoxamine, which removes more iron than each one of the drugs by itself.

Deferasirox (ICL670), another oral iron chelator, is particularly promising for its efficacy, which may be similar to that of deferoxamine. Deferasirox is administered once daily and appears to have an acceptable side-effect profile.

Bone marrow transplantation had excellent results in low-risk patients, those with thalassemia termed class 1 or class 2 by the Lucarelli classification, which is used to assess risk factors that predict outcome and prognosis and addresses the degree of hepatomegaly, the presence of portal fibrosis on liver biopsy, and the effectiveness of chelation therapy before transplantation. However, patients with class 3 disease (with extensive liver damage from iron overload) have had poor results in the past, primarily because of the 30 percent rate of graft rejection due to attenuated conditioning.

The use of related or unrelated umbilical-cord blood further increases the donor pool. However, cordblood transplantations have often been unsuccessful in the treatment of thalassemia because large numbers of transplanted cells need to be administered to sustain hematopoiesis and prevent graft rejection.

Initial efforts at gene therapy were directed against diseases of the β -globin gene. This therapeutic strategy involves the insertion of a normally functioning γ -globin or β -globin gene into the patient's autologous hematopoietic stem cells. Although the concept is relatively straightforward, it has met with two decades of seemingly insurmountable hurdles and is still in experimental stage.

Polymerase-chain-reaction (PCR) technology has been used for more than a decade for prenatal diagnosis to detect point mutations or deletions in chorionic-villus samples, enabling first-trimester, DNA-based testing for thalassemia. However, because pregnancy termination is unacceptable to some persons (even when the fetus is affected), methods were developed beginning in the early 1990s, to perform diagnostic testing before implantation, but they are still not commonly used.

In developed parts of the world, such as the United States and Europe, there are approximately 10,000 homozygous patients with thalassemia, and the number of new cases has been progressively decreasing because of effective prevention methods.

In contrast, the treatment of thalassemia is entirely different in less developed countries, where most of the patients with this disease reside. Safe transfusion (with the use of filtration and the viral testing of blood) and chelation are not universally available. Consequently, many patients with thalassemia in underdeveloped nations die in childhood or adolescence.

The challenge for the future is to ensure that people who are born with a severe form of thalassemia will continue to thrive, while effective prevention eventually decreases the number of severely affected patients worldwide.

β-THALASSEMIA

Thalassemia is a hereditary anemia resulting from defects in hemoglobin production.1 β -Thalassemia, which is caused by a decrease in the production of β -globin chains (Fig. 1), affects multiple organs and is associated with considerable morbidity and mortality. 2 Accordingly, lifelong care is required,3 and financial expenditures for proper treatment are substantial.4

Thalassemia is among the most common genetic disorders worldwide; 4.83 percent of the world's population carry globin variants, including 1.67 percent of the population who are heterozygous for α -thalassemia and β -thalassemia. In addition, 1.92 percent carry sickle hemoglobin, 0.95 percent carry hemoglobin E, and 0.29 percent carry hemoglobin C. Thus, the worldwide birth rate of people who are homozygous or compound heterozygous for symptomatic globin disorders, including α -thalassemia and β -thalassemia, is no less than 2.4 per 1000 births, of which 1.96 have sickle cell disease and 0.44 have thalassemias.5

REFERENCES

- Higgs DR, Thein SL, Woods WG. The molecular pathology of the thalassaemias. In: Weatherall DJ, Clegg B, eds. The thalassaemia syndromes. 4th ed. Oxford, England: Blackwell Science, 2001:133-91.
- 2. Cunningham MJ, Macklin EA, Neufeld EJ, Cohen AR.

- Complications of b-thalassemia major in North America. Blood 2004: 104:34-9
- Old JM, Olivieri NF, Thein SL. Diagnosis and management of thalassaemia. In: Weatherall DJ, Clegg B, eds. The thalassaemia syndromes. 4th ed. Oxford, England: Blackwell Science, 2001:630-85.
- Karnon J, Zeuner D, Brown J, Ades AE, Wonke B, Modell B. Lifetime treatment costs of b-thalassaemia major. Clin Lab Haematol 1999;21:377-85.
- Angastiniotis M, Modell B. Global epidemiology of hemoglobin disorders. Ann N Y Acad Sci 1998;850:251-9.
- Weatherall DJ. Phenotype-genotype relationships in monogenic disease: lessons from the thalassaemias. Nat Rev Genet 2001; 2:245-55.
- Kong Y, Zhou S, Kihm AJ, et al. Loss of alpha-hemoglobinstabilizing protein impairs erythropoiesis and exacerbates bthalassemia. J Clin Invest 2004;114:1457-66.
- Pootrakul P, Sirankapracha P, Hemsorach S, et al. A correlation of erythrokinetics, ineffective erythropoiesis, and erythroid precursor apoptosis in Thai patients with thalassemia. Blood 2000;96:2606-12.
- Rachmilewitz EA, Schrier S. The pathophysiology of bthalassemia. In: Steinberg MH, Forget BG, Higgs DR, Nagel RL, eds. Disorders of hemoglobin: genetics, pathophysiology, and clinical management. Cambridge, England: Cambridge University Press, 2001:233-51.
- Advani R, Sorenson S, Shinar E, Lande W, Rachmilewitz E, Schrier SL. Characterization and comparison of the red blood cell membrane damage in severe human a- and b-thalassemia. Blood 1992;79:1058-63.
- Higgs DR, Thein SL, Wood WG. The pathophysiology of the thalassaemias. In: Weatherall DJ, Clegg B, eds. The thalassaemia syndromes. 4th ed. Oxford, England: Blackwell Science, 2001:192-236.
- Shinar E, Rachmilewitz EA, Lux SE. Differing erythrocyte membrane skeletal protein defects in alpha and beta thalassemia. J Clin Invest 1989;83:404-10.
- Hershko C, Graham G, Bates GW, Rachmilewitz EA. Nonspecific serum iron in thalassemia: an abnormal serum iron fraction of potential toxicity. Br J Haematol 1978;40: 255-63.
- Kuypers FA, de Jong K. The role of phosphatidylserine in recognition and removal of erythrocytes. Cell Mol Biol 2004;50:147-58.
- Tavazzi D, Duca L, Graziadei G, Comino A, Fiorelli G, Cappellini MD. Membranebound iron contributes to oxidative damage of b-thalassemia intermedia erythrocytes. Br J Haematol 2001;112:48-50.
- Eldor A, Rachmilewitz EA. The hypercoagulable state in thalassemia. Blood 2002; 99:36-43.
- Centis F, Tabellini L, Lucarelli G, et al. The importance of erythroid expansion in determining the extent of apoptosis in erythroid precursors in patients with b-thalassemia major. Blood 2000;96:3624-9.
- Mathias LA, Fisher TC, Zeng L, et al. Ineffective erythropoiesis in b-thalassemia major is due to apoptosis at the polychro matophilic normoblast stage. Exp Hematol 2000;28:1343-53.
- De Maria R, Testa U, Luchetti L, et al. Apoptotic role of Fas/ Fas ligand system in the regulation of erythropoiesis. Blood 1999;93:796-803.
- Testa U. Apoptotic mechanisms in the control of erythropoiesis.
 Leukemia 2004; 18:1176-99.
- Angelucci E, Bai H, Centis F, et al. Enhanced macrophagic attack on b-thalassemia major erythroid precursors. Haematologica 2002;87:578-83.
- Dzik WH. Leukoreduction of blood components. Curr Opin Hematol 2002;9: 521-6.
- De Sanctis V. Growth and puberty and its management in thalassaemia. Horm Res 2002;58:Suppl 1:72-9.

- Raiola G, Galati MC, De Sanctis V, et al. Growth and puberty in thalassemia major. J Pediatr Endocrinol Metab 2003;16:Suppl 2:259-66
- Cohen AR, Galanello R, Pennell DJ, Cunningham MJ, Vichinsky E. Thalassemia. Hematology (Am Soc Hematol Educ Program) 2004;1:14-34.
- Voskaridou E, Terpos E. New insights into the pathophysiology and management of osteoporosis in patients with beta thalassaemia. Br J Haematol 2004;127:127-39.
- Modell B, Khan M, Darlison M. Survival in beta-thalassaemia major in the UK: data from the UK Thalassaemia Register. Lancet 2000;355:2051-2.
- 28. Olivieri NF. The b-thalassemias. **N Engl J Med** 1999;341:99-109. [Erratum, N Engl J Med 1999;341:1407.]
- Ganz T. Hepcidin, a key regulator of iron metabolism and mediator of anemia of inflammation. Blood 2003;102:783-8.
- Papanikolaou G, Tzilianos M, Christakis Π, et al. Hepcidin in iron overload disorders. Blood 2005;105:4103-5.
- Rachmilewitz EA, Weizer-Stern O, Adamsky K, et al. Iron and oxidative stress in thalassemia: Eighth International Cooley's Anemia Symposium, March 2005. Ann NY Acad Sci 2005;1054:1-6
- Hoffbrand AV, Cohen A, Hershko C. Role of deferiprone in chelation therapy for transfusional iron overload. Blood 2003;102: 17-24.
- Brittenham GM, Cohen AR, McLaren CE, et al. Hepatic iron stores and plasma ferritin concentration in patients with sickle cell anemia and thalassemia major. Am J Hematol 1993;42:81-5.
- 34. Anderson LJ, Wonke B, Prescott E, Holden S, Walker JM, Pennell DJ. Comparison of effects of oral deferiprone and subcutaneous desferrioxamine on myocardial iron concentrations and ventricular function in beta-thalassemia. Lancet 2002;360: 516-20.
- Voskaridou E, Douskou M, Terpos E, et al. Magnetic resonance imaging in the evaluation of iron overload in patients with b thalassemia and sickle cell disease. Br J Haematol 2004;126:736-42.
- Esposito BP, Breuer W, Sirankapracha P, Pootrakul P, Hershko C, Cabantchik ZI. Labile plasma iron in iron overload: redox activity and susceptibility to chelation. Blood 2003; 102:2670-7.
- Borgna-Pignatti C, Rugolotto S, De Stefano P, et al. Survival and complications in patients with thalassemia major treated with transfusion and deferoxamine. Haematologica 2004;89:1187-93.
- Franchini M, Veneri D. Iron-chelation therapy: an update. Hematol J 2004;5:287-92.
- Wanless IR, Sweeney G, Dhillon AP, et al. Lack of progressive hepatic fibrosis during long-term therapy with deferiprone in subjects with transfusion-dependent b-thalassemia. Blood 2002;100:1566-9. [Erratum, Blood 2003;101:2460.]
- Ceci A, Baiardi P, Felisi M, et al. The safety and effectiveness of deferiprone in a large-scale, 3-year study in Italian patients. Br J Haematol 2002;118:330-6.
- Shalev O, Repka T, Goldfarb A, et al. Deferiprone (L1) chelates pathologic iron deposits from membranes of intact thalassemic and sickle red blood cells both in vitro and in vivo. Blood 1995;86:2008-13.
- Pootrakul P, Sirankapracha P, Sankote J, et al. Clinical trial of deferiprone iron chelation therapy in b-thalassaemia/haemoglobin E patients in Thailand. Br J Haematol 2003;122:305-10.
- 43. Piga A, Gaglioti C, Fogliacco E, Tricta F. Comparative effects of deferiprone and deferoxamine on survival and cardiac disease in patients with thalassemia major: a retrospective analysis. **Haematologica** 2003;88: 489-96.
- 44. Hershko C, Link GM, Konijn AM, Cabantchik ZI. Iron chelation therapy. Curr Hematol Rep 2005;4:110-6.
- Giardina PJ, Grady RW. Chelation therapy in b thalassemia: an optimistic update. Semin Hematol 2001;38:360-6.
- Link G, Konijn AM, Breuer W, Cabantchik ZI, Hershko C. Exploring the "iron shuttle" hypothesis in chelation therapy:

- effects of combined deferoxamine and deferiprone treatment in hypertransfused rats with labeled iron stores and in iron-loaded rat heart cells in culture. **J Lab Clin Med** 2001; 138:130-8.
- 47. Mourad FH, Hoffbrand AV, Sheikh-Taha M, Koussa S, Khoriaty AI, Taher A. Comparison between desferrioxamine and combined therapy with desferrioxamine and deferiprone in iron overloaded thalassaemia patients. Br J Haematol 2003;121:187-9.
- 48. Cappellini M, Bejaoui M, Perrotta S, et al. Phase III evaluation of once-daily, oral therapy with ICL670 (Exjade) versus deferoxamine in patients with b-thalassemia and transfusional hemosiderosis. Blood 2004; 104:984a. abstract.
- 49. Galanello R, Piga A, Alberti D, Rouan MC, Bigler H, Sechaud R. Safety, tolerability, and pharmacokinetics of ICL670, a new orally active iron-chelating agent in patients with transfusion-dependent iron overload due to b-thalassemia. J Clin Pharmacol 2003; 43:565-72.
- Nisbet-Brown E, Olivieri NF, Giardina PJ, et al. Effectiveness and safety of ICL670 in iron-loaded patients with thalassaemia: a randomised, double-blind, placebo-controlled, dose-escalation trial. Lancet 2003; 361:1597-602.
- Cappellini MD, Robbiolo L, Bottasso BM, Coppola R, Fiorelli G, Mannucci AP. Venous thromboembolism and hypercoagulability in splenectomized patients with thalassaemia intermedia. Br J Haematol 2000;111:467-73.
- Manodori AB, Barabino GA, Lubin BH, Kuypers FA. Adherence of phosphatidylserine- exposing erythrocytes to endothelial matrix thrombospondin. Blood 2000;95: 1293-300.
- Amer J, Fibach E. Oxidative status of platelets in normal and thalassemic blood. Thromb Haemost 2004;92:1052-9.
- Lucarelli G, Clift RA, Galimberti M, et al. Marrow transplantation for patients with thalassemia: results in class 3 patients. Blood 1996;87:2082-8.
- Sodani P, Gaziev D, Polchi P, et al. New approach for bone marrow transplantation in patients with class 3 thalassemia aged younger than 17 years. Blood 2004; 104:1201-3.
- Iannone R, Casella JF, Fuchs EJ, et al. Results of minimally toxic nonmyeloablative transplantation in patients with sickle cell anemia and beta-thalassemia. Biol Blood Marrow Transplant 2003;9:519-28.
- 57. Hongeng S, Chuansumrit A, Hathirat P, Rerkamnuaychoke B, Chaisiripoomkere W, Jootar S. Full chimerism in nonmyeloablative stem cell transplantation in a beta-thalassemia major patient (class 3 Lucarelli). Bone Marrow Transplant 2002;30:409-10.
- Andreani M, Nesci S, Lucarelli G, et al. Long-term survival of exthalassemic patients with persistent mixed chimerism after bone marrow transplantation. Bone Marrow Transplant 2000;25:401-4.
- Krishnamurti L, Abel S, Maiers M, Flesch S. Availability of unrelated donors for hematopoietic stem cell transplantation for hemoglobinopathies. Bone Marrow Transplant 2003;31:547-50.
- La Nasa G, Giardini C, Argiolu F, et al. Unrelated donor bone marrow transplantation for thalassemia: the effect of extended haplotypes. Blood 2002;99:4350-6.
- Locatelli F, Rocha V, Reed W, et al. Related umbilical cord blood transplantation in patients with thalassemia and sickle cell disease. Blood 2003;101:2137-43.
- Fang J, Huang S, Chen C, et al. Umbilical cord blood transplantation in Chinese children with beta-thalassemia. J Pediatr Hematol Oncol 2004;26:185-9.
- Hall JG, Martin PL, Wood S, Kurtzberg J. Unrelated umbilical cord blood transplantation for an infant with beta-thalassemia major. J Pediatr Hematol Oncol 2004;26:382-5.
- 64. Tan PH, Hwang WY, Goh YT, et al. Unrelated peripheral blood and cord blood hematopoietic stem cell transplants for thalassemia major. Am J Hematol 2004;75:209-12.
- Li CK, Chik KW, Wong GW, Cheng PS, Lee V, Shing MM. Growth and endocrine function following bone marrow transplantation for thalassemia major. Pediatr Hematol Oncol 2004;21:411-9.
- 66. Atweh GF, Loukopoulos D. Pharmacological induction of fetal

- hemoglobin in sickle cell disease and b-thalassemia. **Semin Hematol** 2001;38:367-73.
- 67. Steinberg MH, Barton F, Castro O, et al. Effect of hydroxyurea on mortality and morbidity in adult sickle cell anemia: risks and benefits up to 9 years of treatment. JAMA 2003;289:1645-51. [Erratum, JAMA 2003; 290:756.]
- Bradai M, Abad MT, Pissard S, Lamraoui F, Skopinski L, de Montalembert M. Hydroxyurea can eliminate transfusion requirements in children with severe b-thalassemia. Blood 2003;102:1529-30.
- Gilman JG, Huisman TH. DNA sequence variation associated with elevated fetal G gamma globin production. Blood 1985;66: 783-7.
- Fucharoen S, Siritanaratkul N, Winichagoon P, et al. Hydroxyurea increases hemoglobin F levels and improves the effectiveness of erythropoiesis in b-thalassemia/hemoglobin E disease. Blood 1996;87:887-92.
- Rachmilewitz EA, Aker M, Perry D, Dover G. Sustained increase in haemoglobin and RBC following long-term administration of recombinant human erythropoietin to patients with homozygous b-thalassaemia. Br J Haematol 1995;90:341-5.
- 72. Singer ST, Sweeters N, Vichinsky EP, Wagner AJ, Rachmilewitz EA. A dose-finding and safety study of darbepoietin alfa (erythropoiesis stimulating protein) for the treatment of anemia in patients with thalassemia intermedia. Blood 2003;102:268a. abstract.
- 73. Fibach E. Cell culture and animal models to screen for promising fetal hemoglobin- stimulating compounds. Semin Hematol 2001;38:374-81. 74. Bhanu NV, Trice TA, Lee YT, Miller JL. A signaling mechanism for growth-related expression of fetal hemoglobin. Blood 2004; 103:1929-33.
- 75. Cighetti G, Duca L, Bortone L, et al. Oxidative status and malondialdehyde in b-thalassemia patients. Eur J Clin Invest 2002;32:Suppl 1:55-60. 76. Tesoriere L, D'Arpa D, Butera D, et al. Oral supplements of vitamin E improve measures of oxidative stress in plasma and reduce oxidative damage to LDL and erythrocytes in b-thalassemia intermedia patients. Free Radic Res 2001;34:529-40.
- Rund D, Rachmilewitz EA. Pathophysiology of a- and b-thalassemia: therapeutic implications. Semin Hematol 2001;38:343-9.
- Pace BS, Shartava A, Pack-Mabien A, Mulekar M, Ardia A, Goodman SR. Effects of N-acetylcysteine on dense cell formation in sickle cell disease. Am J Hematol 2003; 73:26-32.
- Persons DA, Nienhuis AW. Gene therapy for the hemoglobin disorders. Curr Hematol Rep 2003;2:348-55.
- Puthenveetil G, Malik P. Gene therapy for hemoglobinopathies: are we there yet? Curr Hematol Rep 2004;3:298-305.
- May C, Rivella S, Callegari J, et al. Therapeutic haemoglobin synthesis in betathalassaemic mice expressing lentivirusencoded human beta-globin. Nature 2000; 406:82-6.
- Rivella S, Sadelain M. Therapeutic globin gene delivery using lentiviral vectors. Curr Opin Mol Ther 2002;4:505-14.
- 83. Moreau-Gaudry F, Xia P, Jiang G, et al. High-level erythroidspecific gene expression in primary human and murine hematopoietic cells with self-inactivating lentiviral vectors. **Blood** 2001;98:2664-72.
- 84. Hanawa H, Hargrove PW, Kepes S, Srivastava DK, Nienhuis AW, Persons DA. Extended beta-globin locus control region elements promote consistent therapeutic expression of a gamma-globin lentiviral vector in murine beta-thalassemia. Blood 2004; 104:2281-90.
- 85. Lung HY, Meeus IS, Weinberg RS, Atweh GF. In vivo silencing of the human gammaglobin gene in murine erythroid cells following retroviral transduction. **Blood Cells Mol Dis** 2000;26:613-9.
- Rivella S, Callegari JA, May C, Tan CW, Sadelain M. The cHS4 insulator increases the probability of retroviral expression at random chromosomal integration sites. J Virol 2000;74:4679-87.

- 87. Emery DW, Yannaki E, Tubb J, Nishino T, Li Q, Stamatoyanno poulos G. Development of virus vectors for gene therapy of beta chain hemoglobino pathies: flanking with a chromatin insulator reduces gammaglobin gene silencing in vivo. **Blood** 2002; 100:2012-9.
- Puthenveetil G, Scholes J, Carbonell D, et al. Successful correction of the human beta-thalassemia major phenotype using a lentiviral vector. Blood 2004;104:3445-53.
- Imren S, Fabry ME, Westerman KA, et al. High-level beta-globin expression and preferred intragenic integration after lentiviral transduction of human cord blood stem cells. J Clin Invest 2004;114:953-62.
- Zoueva OP, Rodgers GP. Inhibition of beta protein 1 expression enhances betaglobin promoter activity and beta-globin mRNA levels in the human erythroleukemia (K562) cell line. Exp Hematol 2004;32:700-8.
- Lacerra G, Sierakowska H, Carestia C, et al. Restoration of hemoglobin A synthesis in erythroid cells from peripheral blood of thalassemic patients. Proc Natl Acad Sci U S A 2000;97:9591-6.
- Vacek MM, Ma H, Gemignani F, Lacerra G, Kafri T, Kole R. High-level expression of hemoglobin A in human thalassemic erythroid progenitor cells following lentiviral vector delivery of an antisense snRNA. Blood 2003;101:104-11.
- Braude P, Pickering S, Flinter F, Ogilvie CM. Preimplantation genetic diagnosis. Nat Rev Genet 2002;3:941-53. [Erratum, Nat Rev Genet 2003;4:157.]
- 94. Piyamongkol W, Bermudez MG, Harper JC, Wells D. Detailed investigation of factors influencing amplification efficiency and

- allele drop-out in single cell PCR: implications for preimplantation genetic diagnosis. **Mol Hum Reprod** 2003;9:411-20.
- Fiorentino F, Biricik A, Karadayi H, et al. Development and clinical application of a strategy for preimplantation genetic diagnosis of single gene disorders combined with HLA matching. Mol Hum Reprod 2004;10: 445-60.
- Van de Velde H, Georgiou I, De Rycke M, et al. Novel universal approach for preimplantation genetic diagnosis of betathalassaemia in combination with HLA matching of embryos. Hum Reprod 2004;19:700-8.
- Di Naro E, Ghezzi F, Vitucci A, et al. Prenatal diagnosis of bthalassemia using fetal erythroblasts enriched from maternal blood by a novel gradient. Mol Hum Reprod 2000; 6:571-4.
- Ding C, Chiu RWK, Lau TK, et al. MS analysis of singlenucleotide differences in circulating nucleic acids: application to noninvasive prenatal diagnosis. Proc Natl Acad Sci U S A 2004;101:10762-7.
- Cao A, Galanello R. Effect of consanguinity on screening for thalassemia. N Engl J Med 2002;347:1200-2. 100. Anie KA, Massaglia P. Psychological therapies for thalassaemia. Cochrane Database Syst Rev 2001;3:CD002890.
- 101. Ahmed S, Saleem M, Modell B, Petrou M. Screening extended families for genetic hemoglobin disorders in Pakistan. N Engl JMed 2002;347:1162-8.
- 102. Alwan A, Modell B. Recommendations for introducing genetics services in developing countries. Nat Rev Genet 2003; 4:61-8. Copyright © 2005 Massachusetts Medical Society.