

# Tratamiento estándar de los síndromes mielodisplásicos adaptados al riesgo

Risk-adapted standard treatment  
of myelodysplastic syndromes

Hellström-Lindberg E, Tobiasson M, Jädersten M

*Center for Hematology and Regenerative Medicine,  
Karolinska Institutet Karolinska University Hospital Stockholm Sweden*

Eva.hellstrom-lindberg@ki.se



SIMPOSIO CONJUNTO  
EHA-SAH:  
SÍNDROMES  
MIELODISPLÁSICOS

HEMATOLOGÍA  
Volumen 21 N° Extraordinario: 316-327  
XXIII Congreso Argentino  
de Hematología  
Noviembre 2017

**Palabras claves:** SMD,  
tratamiento,  
mutaciones.

**Keywords:** MDS,  
treatment,  
mutations.

## Abstract

Treatment of MDS has since many years been based mainly on conventional risk scores dividing patients into lower and higher-risk MDS. During recent years the fast development of knowledge about mutational profiles has added significantly to the decision-making and has also fostered development of novel targeted drugs in myeloid malignancies. Moreover, the possibilities to guide patients through the hitherto only curative therapeutic option, allogeneic stem cell transplantation, have improved markedly. Hence NGS sequencing is today affecting diagnostics, prognostication, and choice of treatment, as well as the tools for detection of minimal residual disease, and will develop to a natural part of future patient management.

## Introduction

The myelodysplastic syndromes (MDS) constitute a heterogeneous group of myeloid malignancies originating in the hematopoietic stem and progenitor cell compartment (HSPC), an established statement that not until recently has been scientifically validated in MDS<sup>(1-3)</sup>.

The cellular origin of the disease, as well as the molecular disease mechanisms leading to specific disease phenotype and patient symptoms is, however, utterly important for the choice of treatment and for the development of novel therapeutics.

Classical low-risk MDS subtypes including 5q-syndrome and MDS with ring sideroblasts (MDS-RS, according to WHO 2016 classification<sup>(4)</sup>) arise in rare phenotypically intact multipotent hematopoietic

stem cells (HSC).

These initiating cancer stem cells can subsequently be subject to acquisition of additional mutations leading to altered disease phenotype including disease progression. Interestingly, the risk of clonal evolution seems to be more frequent in del(5q) MDS<sup>(5)</sup> than in MDS-RS and some other low-risk subtypes, which needs to be considered when long-term treatment is planned, including evaluation for potentially curative allogeneic stem cell transplantation (SCT)<sup>(5)</sup>.

Higher-risk MDS subtypes are usually characterized by compromised HPSC phenotypes, as recently shown in a study of MDS with cytogenetic alterations encompassing chromosome 7<sup>(3)</sup>. In these patients the clonal expansion, similar to what has been shown in acute myeloid leukemia (AML), usually arises in other subsets within the HPSC compartment.

The propensity for clonal instability and evolution is generally higher than in lower-risk MDS. Both low and high-risk MDS are considered incurable by conventional chemotherapy or other currently available therapies than allogeneic stem cell transplantation.

### Standard risk assessment

Patients with MDS have an overall poor outcome, however, with a considerable variation between patient subgroups. The standard risk tool for patients with MDS is since 2012 the revised International Prognostic Scoring System (IPSS-R), dividing patients into 5 risk groups according to the degree of anemia, neutropenia, and thrombocytopenia, bone marrow blast percentage, and cytogenetic risk profile (Table 1)<sup>(6)</sup>. The IPSS-R has been validated in several publications, including its ability to guide decision-making for example with regard to SCT<sup>(7,8)</sup>. One important aspect is that a diagnosis of MDS always implies a decreased relative survival, i.e. a shorter life expectancy than for an age-and sex-matched person without MDS. In addition to the IPSS-R score, prognosis is influenced by the presence of grade 2 or 3 fibrosis which is independently associated with inferior outcome<sup>(9)</sup>. Moreover, comorbidities are also important, in particular when planning for more toxic treatment modalities. An MDS-specific index (MDS-CI) has been developed based on presence of: cardiac, liver, renal, or pulmonary disease, or presence of other tumors<sup>(10)</sup>.

**Table 1**

| Risk factor  | 0         | +0.5  | +1        | +1.5 | +2           | +4   | +5        |
|--------------|-----------|-------|-----------|------|--------------|------|-----------|
| Cytogenetics | Very good | -     | Good      | -    | Intermediate | Poor | Very poor |
| BM blasts, % | ≤2%       | -     | 2.1%-4.9% | -    | 5%-10%       | >10% | -         |
| Hemoglobin   | ≥100      | -     | 80-99     | <80  |              |      |           |
| Platelets    | ≥100      | 50-99 | <50       |      |              |      |           |
| ANC          | ≥0.8      | <0.8  |           |      |              |      |           |

| Risk group   | Risk score | Patient distribution (%) | Median OS (years) | Median time to 25% AML evolution (years) |
|--------------|------------|--------------------------|-------------------|--|
| Very low     | ≤1.5       | 19                       | 8.8               | Not reached                              |
| Low          | 2-3        | 38                       | 5.3               | 10.8                                     |
| Intermediate | 3.5-4.5    | 20                       | 3.0               | 3.2                                      |
| High         | 5-6        | 13                       | 1.6               | 1.4                                      |
| Very high    | >6         | 10                       | 0.8               | 0.7                                      |

#### Cytogenetic group

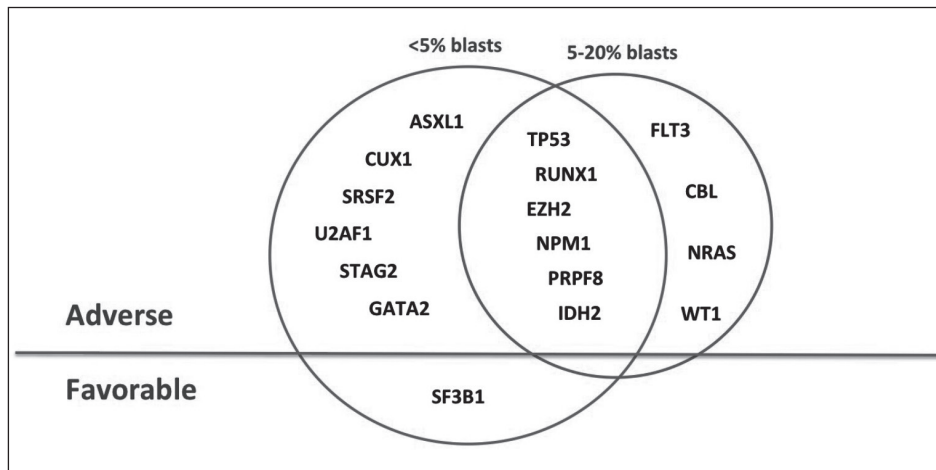
- **Very good:** del(11q) or -Y
- **Good:** normal karyotype, del(20q), del(5q), del(12p), or double including del(5q)
- **Intermediate:** +8, del(7q), i(17q), +19, or any other single or double independent clone
- **Poor:** -7, inv(3)/t(3q)/del(3q), double including -7/del(7q), or complex (3 abnormalities)
- **Very poor:** complex >3 abnormalities

### Molecular diagnostics and additional prognostic factors

Mutation screening has in recent years become readily available and more affordable. The presence of adverse mutations may support the clinical decision to transplant a low or intermediate risk patient with MDS, in particular in case of severe cytopenias. Both the type and number of mutations have prognostic implications<sup>(11-14)</sup>. Mutations associated with poor prognosis include: TP53, EZH2, ETV6, RUNX1, NRAS, and ASXL1. The only mutation consistently associated with favorable outcome is SF3B1, commonly observed in MDS-RS. AML-like mutations such as NPM1, NRAS, FLT3 are associated with transformed disease.

Aside from prognostic implications, mutational screening can be of value in borderline cases where a morphologic diagnosis of MDS is challenging; 80-90% of MDS patients carry at least one recurrent mutation<sup>(11,12)</sup>. However, a caveat is that the aging bone marrow is increasingly prone to harbor clonal hematopoiesis, without necessarily resulting in cytopenias or disease. The term CHIP (clonal hematopoiesis of indeterminate potential) is used for this condition, and it carries approximately a 1% annual risk of evolving to MDS<sup>(15)</sup>.

Currently, a rational approach could be to genetically characterize transplant candidates as well as unexplained cytopenias that do not fulfill the criteria of MDS. (**Figura 1**)



**Figure 1.** Most mutations may occur at early phases of MDS and can cooperate to cause disease progression. A few mutations are tightly associated with blast increase, and only SF3B1 is linked to favorable outcome.

### Treatment of MDS

Patients with MDS are treated of two main reasons: To prolong survival and hopefully cure the disease, and to improve symptoms, thereby quality of life. Naturally these two purposes frequently overlap. In the Nordic guidelines for MDS, we have stressed the process that all newly diagnosed patients below the age of 75 years should undergo molecular risk profiling and be evaluated for a potential cure with stem cell transplantation. Still, a majority of these patients will probably not proceed to SCT, but we know for certain that transplantation after further clonal evolution and progression to overt high-risk MDS or acute leukemia is significantly associated with decreased outcome after transplantation.

### TREATMENT WITH ERYTHROPOIESIS-STIMULATING AGENTS (ESAs)

Anemia is often the only significant cytopenia in lower-risk MDS. The anemia of MDS and chronic transfusion-dependency are according to most studies significantly associated with reduced quality of life and decreased survival<sup>(7,16)</sup>. When comparing patients with and without transfusion need but with the same IPSS-R score, transfusion dependency is significantly associated with both reduced survival and a higher risk for disease progression. The reason for the poorer outcome is probably multifactorial and includes effects of chronic anemia on cardiovascular morbidity and chronic iron overload. It is also likely that transfusion need reflects more severe disease biology.

Erythropoietin (EPO) is first-line treatment for the anemia of lower-risk MDS, according to the European guidelines and should be offered to patients with symptomatic anemia<sup>(17)</sup>. The efficacy of EPO may be enhanced or restored by the addition of granulocyte-CSF (G-CSF), which acts synergistically with EPO to inhibit mitochondria-mediated erythroid apoptosis. Based on studies performed more than 20 years ago when patients usually were treated later in their disease, a transfusion need equal or above 2 red blood cell units per month and a serum erythropoietin  $\geq 500$  U/l predicted for a very poor response to treatment. This is still valid and ESAs should not be used in these patients, often characterized by marked erythroid hypoplasia in the bone marrow. However, as more treatment options emerge for patients with lower-risk MDS, better tools are needed to discriminate patients with poor and intermediate response probabilities, respectively. In a recent study, Buckstein and colleagues describe 996 ESA-treated patients from three registries<sup>(18)</sup>. Overall response rate was 59%, which reflects the fact that patients today are treated earlier in their disease. By multivariate analysis, transfusion independence, erythropoietin (EPO) level  $< 100$  IU/L, and IPSS low-risk were independently predictive of response. Assigning a score of 1 to each resulted in a scoring system of 0-3 with response rates of 23%, 43%, 67%, and 85%. This new 'ITACA' score has a higher discriminating power of response than previously published scores.

ESA treatment has been associated with an improved survival in large retrospective studies mainly encompassing studies performed 1990 to 2000, using matched untreated patient cohorts as control<sup>(12,13,19,20)</sup>. The EU MDS Registry prospectively enrolls patients with lower-risk MDS treated within routine clinical practice at smaller and larger European hospitals. A recent study describes the outcome of 1696 patients in the registry, randomly treated with ESA or not based on local routines, reimbursement policy and other factors. Importantly, ESA treatment initiated in patients with Hb levels less than 10 g/dl but prior to a permanent transfusion need significantly delayed the onset of chronic transfusion dependency ( $p < 0.0001$ ). These recent large studies clearly shows that early ESA treatment is the preferred mode of treatment, which should be reflected in future guidelines and National reimbursement policies.

#### IMMUNOSUPPRESSIVE TREATMENT IN LOWER-RISK MDS

A small fraction of low risk MDS patients with MDS-SLD and MDS-MLD seem to have bone marrow failure due to autoimmune mechanisms, as known from aplastic anemia<sup>(21)</sup>. Several international studies have demonstrated response rates in the order of 30% to immunosuppressive therapy (antithymocyte globulin [ATG] in some investigations combined with cyclosporin A [CyA]) in patients with MDS-SLD and MDS-MLD. HLA-DR15 positivity, young age and short duration of red cell transfusion dependence seem to predict for a response to immunosuppressive therapy in MDS patients, although this is based on a limited material. An analysis of patients treated at NIH indicated an improvement in survival of ATG treated patients, especially in younger individuals with lower risk disease. To date, there are no controlled data to support the addition of cyclosporin A to ATG treatment in MDS, although this combination has been shown to increase the response rate in a retrospective analysis.

#### NOVEL THERAPEUTIC OPTIONS FOR MDS WITH RING SIDEROBLASTS

Luspatercept has emerged as a novel treatment for EPO resistant anemia in MDS with ring sideroblasts, as recently reported in a pivotal Phase I-II study showing IWG hematologic improvement of better in 63% of 51 patients treated with effective doses<sup>(22)</sup>. 42 patients were evaluable for transfusion independency and 16 of these (38%) achieved RBC-TI. Of the 22 patients who entered the extension phase study, 50% had a continued response with a median duration of 15 months. Treatment response was significantly associated with mutations in SF3B1, a core component of the spliceosome. Hence, novel therapeutic alternatives targeted to specific molecular alterations are part of the future for treatment of lower-risk MDS.

#### MDS del(5q) and lenalidomide

Lenalidomide binds to cereblon, a substrate adaptor of the CRL4<sup>CRBN</sup> E3 ubiquitin ligase, and modulates the ubiquitination and degradation of specific proteins. One important target is CK1 $\alpha$ , which is already haploinsufficient in MDS with del(5q), and its degradation leads to p53 dependent apoptosis in the MDS cells<sup>(23)</sup>. Lenalidomide is highly efficient in low risk MDS with del(5q), where 43-56% achieve transfusion-independency and 23-57%

show cytogenetic response<sup>(24,25)</sup>. The response rate is better with 10 mg/day 21/28 days compared to 5 mg continuous dosing, without added toxicity. Grade III-IV neutropenia and thrombocytopenia occurs in around 50% of patients, in particular early on during the treatment. The response duration is around 2 years.

The cumulative incidence of AML evolution in treated patients is around 35% at 5 years. Subclones with mutations in TP53 or RUNX1 are tightly linked to disease progression. The mutated clones may be small, down to 1%, and require deep sequencing to be detected. Immunohistochemistry demonstrating strong nuclear staining of p53 is tightly linked to mutation in TP53, and can be used as a surrogate marker<sup>(5,26,27)</sup>. Candidates for allogeneic stem cell transplantations should likely not be treated with lenalidomide; if treatment is given the patient should be carefully monitored for signs of progression. Patients with mutations in TP53 or RUNX1 should be evaluated for alternative treatments due to their adverse prognosis, and lenalidomide should only be considered were no suitable alternative is available.

### Azacitidine

Due to the epigenetic effects of Aza and Dec, these drugs were termed hypomethylating agents. Two large randomized studies have evaluated the effects of Aza on MDS. The first study included all subtypes of MDS (n=99) and patients were randomized to receive either Aza or supportive care. Overall response rate was 60% in the Aza group, which was significantly better than the control arm (p<0.001). There was also a significant difference in progression-free survival (21 vs 12 months; p=0.007) but no significant difference in overall survival could be demonstrated (20 vs 14 months; p=0.1). The next study included only patients with higher-risk MDS (n=357) and randomized these into either Aza or best available treatment which consisted of intensive chemotherapy, low-dose cytarabine or supportive care. In this study a significant survival benefit could be demonstrated (24.5 vs 15 months; p<0.001). Both randomized studies show that responses are normally not seen before the patient has received ≥3 cycles and best response is often seen several cycles after the initial response. A post-hoc analysis of the European study showed a survival benefit also for patients with stable disease<sup>(28)</sup>.

The effect of azacytidine in lower-risk MDS is less well studied and evidence from randomized studies for this treatment is lacking. Several phase-II studies in mostly transfusion-dependent patients, have demonstrated effect in lower-risk MDS although the response rate seems to be lower than in higher-risk patients and a positive effect on long-term outcome has hitherto not been demonstrated. A large randomized study (<https://clinicaltrials.gov/ct2/show/NCT01566695>) assessing the effect of oral Aza in lower-risk patients is ongoing and will hopefully bring more clarity into its role in lower-risk MDS.

Since the response is often delayed several months after start of treatment, predictive tools are highly warranted. Basic clinical data such as morphology and cytogenetics give sparse predictive information, although blast count > 15%, extensive transfusion requirements, abnormal karyotype and previous therapy with cytarabine have been reported as negative predictors of response<sup>(29-31)</sup>. Results from different studies regarding the effects of the mutational profile on response and survival after start of treatment are conflicting. Several studies report higher response rates for *TET2*-mutated patients but the presence of this mutation has not been associated with prolonged survival during treatment<sup>(31-33)</sup>. Another study demonstrated prolonged survival for patients having mutations in any of the histone modulating enzymes (*ASXL1*, *EZH2*)<sup>(34)</sup>. Survival benefit has also been demonstrated for patients with *IDH1/2* mutations (Dec)<sup>(35)</sup>. One study reports high response rates of Dec in TP53-mutated patients with MDS and AML<sup>(36)</sup>. This is however in contrast to other studies on hypomethylating agents on TP53-patients where response and survival is rather reduced<sup>(34,37)</sup>.

An initial reduction of methylation levels after the first treatment cycle in specific genes or on a global level was shown to predict a later clinical response<sup>(38-42)</sup>. If this is reflecting a causative or confounding effect of Aza is however not known. A few studies have reported correlation between methylation level of specific genes e.g. p15 and responses<sup>(43,44)</sup>. In contrast, other studies report no correlation between baseline methylation levels and response<sup>(40,45)</sup>. A recent study has shown that patients with hematopoietic progenitor cells in cell cycle have better response to Aza<sup>(46)</sup>. In summary, larger studies including basic clinical data, mutational and

epigenetic profile is needed to create a clinically useful predictive tool.

The pharmacodynamic effects of Aza are still quite enigmatic. The ruling paradigm for the mechanism of action of Aza claims that promotor demethylation results in re-expression of tumor suppressor genes silenced by aberrant DNA methylation<sup>(40)</sup>. There is however still a lack of convincing evidence confirming this paradigm. In addition to the demethylating effects, there is support for a direct cytotoxic effect, differentiation-promoting effect and direct effects on RNA<sup>(47-49)</sup>. Very interesting in vitro-studies have shown that Aza upregulates retrotransposons which evokes the innate immune system<sup>(50-52)</sup>.

### Other emerging treatments

Several promising therapies are being evaluated in MDS, including drugs targeting specific mutated proteins. Inhibitors of IDH1 and IDH2 are being explored in both MDS and AML, based on highly promising in vitro data. In relapsed/refractory AML, the IDH2 inhibitor enasidenib demonstrated a 40% over all response rate with 19% complete remissions, and the drug was approved for this indication by the FDA in August 2017<sup>(53)</sup>. Preliminary data suggests high efficacy also in MDS, and it will be important to assess whether this is dependent on the clone size or concurrent mutations.

Another target is BCL2, which is an anti-apoptotic protein that is upregulated in high risk MDS and in AML. Preliminary reports of the BCL2 inhibitor venetoclax have demonstrated high activity in MDS and AML both as a single drug and in combination with low-dose Ara-C or hypomethylating agents. In July 2017 it received a breakthrough status for AML by the FDA. Ongoing studies will demonstrate how lasting the responses are, and whether certain subgroups are more sensitive.

Novel hypomethylating agents including guadecitabine and oral azacitidine are being explored both as single drugs as well as in combination with other drugs such as the kinase inhibitor rigosertib. Moreover, early attempts are being made to identify druggable targets caused by alternative splicing, since splicing mutations are seen in around 50% of patients with MDS.

### Allogeneic stem cell transplantation

Allogeneic stem cell transplantation (SCT) for patients with MDS has been performed since the

1970s, initially using sibling donors, but over time with a higher frequency of matched unrelated donors, cord blood or more recently also haploidentical donors. Since allogeneic stem cell transplantation (SCT) is the only potentially curative treatment in MDS, all patients should be evaluated for this option. However, due to the potentially severe complications, transplantation can only be performed in patients up to around 70-75 years of age without significant comorbidity. In younger patients a myeloablative conditioning is normally chosen, while older patients receive a reduced intensity conditioning, which reduces transplantation-related mortality but increases the risk of relapse. Long-term survival rates of between 25% and 45% have been reported after transplantation<sup>(54-56)</sup>. Transplantation-related mortality (TRM) and after myeloablative conditioning and reduced intensity conditioning has been reported to be 32% and 22% and relapse rate 22% vs. 45%, respectively<sup>(54)</sup>. Due to the high risk of TRM, timing of SCT is of great importance where higher-risk patients is recommended to proceed to SCT upfront while lower-risk patients should follow a strict surveillance program and be transplanted in case of signs of progression<sup>(57)</sup>. Recent possibilities to determine what genes are mutated using targeted panel sequencing, enables risk assessment for potential transplantation candidates with higher accuracy and patients with a more indolent disease, as predicted by the risk score, might still be transplantation candidates due to high-risk mutations, e.g. TP53 or RUNX1.

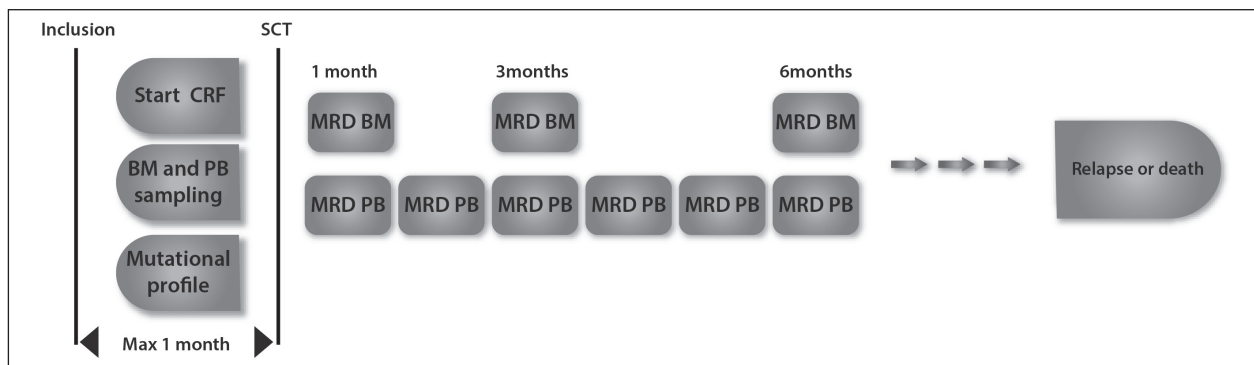
All three prognostic scoring systems (IPSS, IPSS-R and WPSS) have been validated to also predict survival after allogeneic stem cell transplantation<sup>(55,56,58,59)</sup>. The most important risk factor for relapse is genetics. Relapse-free survival at 5 years post transplantation in the five IPSS-R cytogenetic risk groups are reported to be 42%, 36%, 36%, 22% and 10%, respectively<sup>(55)</sup>. More recent data, based on the additional impact of mutations as assessed by targeted panel sequencing, report an increased risk of relapse for patients with mutations in TP53, TET2 or mutations involving RAS-pathway genes<sup>(60,61)</sup>.

In addition to genetics, disease status has impact on survival and debulking treatment e.g. azacitidine or intensive chemotherapy is usually given for patient with a more proliferative disease aiming for the best possible remission prior to transplantation<sup>(17,56)</sup>. The

useful of debulking treatment has however never been tested in prospective controlled studies and there are retrospective studies indicating a similar outcome independent of debulking or not although selection bias is an obvious potential pitfall in these studies<sup>(62,63)</sup>. Likewise, retrospective studies have not been able to demonstrate any advantage for either hypomethylating therapy or intensive chemotherapy as debulking treatment<sup>(64,65)</sup>.

The prognosis after a relapse is dismal although donor lymphocyte infusion might reverse the relapse in rare cases and azacitidine might prolong survival<sup>(66)</sup>. We and others have previously demonstrated that surveillance by using a chimerism analysis can be

used to predict an impending relapse after SCT and serve as trigger for preemptive treatment, however, this analysis has low sensitivity and most often turns positive when the relapse is manifest and no longer possible to treat<sup>(67,68)</sup>. We are in the nordic MDS group now conducting a clinical trial where we evaluate a concept of determining the patient-specific mutations which are then followed by PCR after transplantation systematically in a prospective study (**Figure 2**). Preliminary data indicate that these markers can predict relapse which would enable preemptive treatment before the relapse has become manifest (<https://clinicaltrials.gov/ct2/show/NCT02872662>).



**Figure 2.** Design of the prospective Nordic MDS group study 14B, evaluating patient-specific mutations, followed by PCR as MRD markers after transplantation.

#### Declaración de conflictos de interés:

Martin Jädersten declara haber recibido honorarios por parte de Novartis en concepto de actividades educativas. El resto de los autores declara no poseer conflictos de interés.

#### Bibliografía

1. Woll PS, Kjallquist U, Chowdhury O, Doolittle H, Wedge DC, Thongjuea S, Erlandsson R, Ngara M, Anderson K, Deng Q, Mead AJ, Stenson L, Giustacchini A et al. Myelodysplastic Syndromes Are Propagated by Rare and Distinct Human Cancer Stem Cells In Vivo. *Cancer Cell*. 2014 May 14.
2. Mortera-Blanco T, Dimitriou M, Woll PS, Karimi M, Elvarsdottir E, Conte S, Tobiasson M, Jansson M, Douagi I, Moarii M, Saft L, Papaemmanuil E, Jacobsen SEW et al. SF3B1-initiating mutations in MDS-RSs target lymphomyeloid hematopoietic stem cells. *Blood*. 2017 2017/06/20;130(7):881-90.
3. Dimitriou M, Woll PS, Mortera-Blanco T, Karimi M, Wedge DC, Doolittle H, Douagi I, Papaemmanuil E, Jacobsen SE, Hellstrom-Lindberg E. Perturbed hematopoietic stem and progenitor cell hierarchy in myelodysplastic syndromes patients with monosomy 7 as the sole cytogenetic abnormality. *Oncotarget*. 2016 Nov 08;7(45):72685-98.
4. Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ, Le Beau MM, Bloomfield CD, Cazzola M, Vardiman JW. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood*. 2016 2016/04/11;127(20):2391-405.

5. Scharenberg C, Giai V, Pellagatti A, Saft L, Dimitriou M, Jansson M, Jädersten M, Grandien A, Douagi I, Neuberg DS, LeBlanc K, Boultonwood J, Karimi M et al. Progression in patients with low- and intermediate-1-risk del(5q) myelodysplastic syndromes is predicted by a limited subset of mutations. *Haematologica*. 2016 2016/11/24;102(3):498-508.
6. Greenberg PL, Tuechler H, Schanz J, Sanz G, Garcia-Manero G, Sole F, Bennett JM, Bowen D, Fenaux P, Dreyfus F, Kantarjian H, Kuendgen A, Levis A et al. Revised International Prognostic Scoring System for Myelodysplastic Syndromes. *Blood*. 2012 2012/06/27;120(12):2454-65.
7. de Swart L, Smith A, Johnston TW, Haase D, Droste J, Fenaux P, Symeonidis A, Sanz G, Hellström-Lindberg E, Cermák J, Germing U, Stauder R, Georgescu O et al. Validation of the revised international prognostic scoring system (IPSS-R) in patients with lower-risk myelodysplastic syndromes: a report from the prospective European LeukaemiaNet MDS (EUMDS) registry. *British Journal of Haematology*. 2015 2015/04/24;170(3):372-83.
8. Della Porta MG, Tuechler H, Malcovati L, Schanz J, Sanz G, Garcia-Manero G, Solé F, Bennett JM, Bowen D, Fenaux P, Dreyfus F, Kantarjian H, Kuendgen A et al. Validation of WHO classification-based Prognostic Scoring System (WPSS) for myelodysplastic syndromes and comparison with the revised International Prognostic Scoring System (IPSS-R). A study of the International Working Group for Prognosis in Myelodysplasia (IWG-PM). *Leukemia*. 2015 2015/02/27;29(7):1502-13.
9. Della Porta MG, Malcovati L, Boveri E, Travaglino E, Pietra D, Pascutto C, Passamonti F, Invernizzi R, Castello A, Magrini U, Lazzarino M, Cazzola M. Clinical relevance of bone marrow fibrosis and CD34-positive cell clusters in primary myelodysplastic syndromes. *J Clin Oncol*. 2009 Feb 10;27(5):754-62.
10. Della Porta MG, Malcovati L, Strupp C, Ambaglio I, Kuendgen A, Zipperer E, Travaglino E, Invernizzi R, Pascutto C, Lazzarino M, Germing U, Cazzola M. Risk stratification based on both disease status and extra-hematologic comorbidities in patients with myelodysplastic syndrome. *Haematologica*. 2011 Mar;96(3):441-9.
11. Bejar R, Stevenson K, Abdel-Wahab O, Galili N, Nilsson B, Garcia-Manero G, Kantarjian H, Raza A, Levine RL, Neuberg D, Ebert BL. Clinical effect of point mutations in myelodysplastic syndromes. *N Engl J Med*. 2011 Jun 30;364(26):2496-506.
12. Papaemmanuil E, Gerstung M, Malcovati L, Tauro S, Gundem G, Van Loo P, Yoon CJ, Ellis P, Wedge DC, Pellagatti A, Shlien A, Groves MJ, Forbes SA et al. Clinical and biological implications of driver mutations in myelodysplastic syndromes. *Blood*. 2013 Nov 21;122(22):3616-27; quiz 99.
13. Haferlach T. Molecular genetics in myelodysplastic syndromes. *Leuk Res*. 2012 Dec;36(12):1459-62.
14. Haferlach T, Nagata Y, Grossmann V, Okuno Y, Bacher U, Nagae G, Schnittger S, Sanada M, Kon A, Alpermann T, Yoshida K, Roller A, Nadarajah N et al. Landscape of genetic lesions in 944 patients with myelodysplastic syndromes. *Leukemia*. 2014 Feb;28(2):241-7.
15. Steensma DP, Bejar R, Jaiswal S, Lindsley RC, Sekeres MA, Hasserjian RP, Ebert BL. Clonal hematopoiesis of indeterminate potential and its distinction from myelodysplastic syndromes. *Blood*. 2015 Jul 02;126(1):9-16.
16. Hellstrom-Lindberg E, Gulbrandsen N, Lindberg G, Ahlgren T, Dahl IMS, Dybedal I, Grimfors G, Hesse-Sundin E, Hjorth M, Kanter-Lewensohn L, Linder O, Luthman M, Lofvenberg E et al. A validated decision model for treating the anaemia of myelodysplastic syndromes with erythropoietin + granulocyte colony-stimulating factor: significant effects on quality of life. *British Journal of Haematology*. 2003 2003/03;120(6):1037-46.
17. Malcovati L, Hellstrom-Lindberg E, Bowen D, Ades L, Cermak J, Del Canizo C, Della Porta MG, Fenaux P, Gattermann N, Germing U, Jansen JH, Mittelman M, Mufti G et al. Diagnosis and treatment of primary myelodysplastic syndromes in adults: recommendations from the European LeukemiaNet. *Blood*. 2013 Oct 24;122(17):2943-64.
18. Buckstein R, Balleari E, Wells R, Santini V, Sanna A, Salvetti C, Crisà E, Allione B, Danise P, Finelli C, Clavio M, Poloni A, Salvi F et al. ITACA: A new validated international



- erythropoietic stimulating agent-response score that further refines the predictive power of previous scoring systems. *American Journal of Hematology*. 2017 2017/07/29;92(10):1037-46.
19. Jädersten M, Malcovati L, Dybedal I, Giovanni Della Porta M, Invernizzi R, Montgomery SM, Pascutto C, Porwit A, Cazzola M, Hellström-Lindberg E. Erythropoietin and Granulocyte-Colony Stimulating Factor Treatment Associated With Improved Survival in Myelodysplastic Syndrome. *Journal of Clinical Oncology*. 2008 2008/07/20;26(21):3607-13.
  20. Park S, Grabar S, Kelaidi C, Beyne-Rauzy O, Picard F, Bardet V, Coiteux V, Leroux G, Lepelley P, Daniel MT, Cheze S, Mahe B, Ferrant A et al. Predictive factors of response and survival in myelodysplastic syndrome treated with erythropoietin and G-CSF: the GFM experience. *Blood*. 2008 2008/01/15;111(2):574-82.
  21. Parikh AR, Olnes MJ, Barrett AJ. Immunomodulatory treatment of myelodysplastic syndromes: antithymocyte globulin, cyclosporine, and alemtuzumab. *Semin Hematol*. 2012 Oct;49(4):304-11.
  22. Platzbecker U, Germing U, Gotze KS, Kiewe P, Mayer K, Chromik J, Radsak M, Wolff T, Zhang X, Laadem A, Sherman ML, Attie KM, Giagounidis A. Luspatercept for the treatment of anaemia in patients with lower-risk myelodysplastic syndromes (PACE-MDS): a multicentre, open-label phase 2 dose-finding study with long-term extension study. *Lancet Oncol*. 2017 Sep 01.
  23. Kronke J, Fink EC, Hollenbach PW, MacBeth KJ, Hurst SN, Udeshi ND, Chamberlain PP, Mani DR, Man HW, Gandhi AK, Svinkina T, Schneider RK, McConkey M et al. Lenalidomide induces ubiquitination and degradation of CK1alpha in del(5q) MDS. *Nature*. 2015 Jul 09;523(7559):183-8.
  24. List A, Dewald G, Bennett J, Giagounidis A, Raza A, Feldman E, Powell B, Greenberg P, Thomas D, Stone R, Reeder C, Wride K, Patin J et al. Lenalidomide in the myelodysplastic syndrome with chromosome 5q deletion. *N Engl J Med*. 2006 Oct 05;355(14):1456-65.
  25. Fenaux P, Giagounidis A, Selleslag D, Beyne-Rauzy O, Mufti G, Mittelman M, Muus P, Te Boekhorst P, Sanz G, Del Canizo C, Guerci-Bresler A, Nilsson L, Platzbecker U et al. A randomized phase 3 study of lenalidomide versus placebo in RBC transfusion-dependent patients with Low-/Intermediate-1-risk myelodysplastic syndromes with del5q. *Blood*. 2011 Oct 06;118(14):3765-76.
  26. Jadersten M, Saft L, Smith A, Kulasekararaj A, Pomplun S, Gohring G, Hedlund A, Hast R, Schlegelberger B, Porwit A, Hellstrom-Lindberg E, Mufti GJ. TP53 mutations in low-risk myelodysplastic syndromes with del(5q) predict disease progression. *J Clin Oncol*. 2011 May 20;29(15):1971-9.
  27. Saft L, Karimi M, Ghaderi M, Matolcsy A, Mufti GJ, Kulasekararaj A, Gohring G, Giagounidis A, Selleslag D, Muus P, Sanz G, Mittelman M, Bowen D et al. p53 protein expression independently predicts outcome in patients with lower-risk myelodysplastic syndromes with del(5q). *Haematologica*. 2014 2014/03/28;99(6):1041-9.
  28. Gore SD, Fenaux P, Santini V, Bennett JM, Silverman LR, Seymour JF, Hellstrom-Lindberg E, Swern AS, Beach CL, List AF. A multivariate analysis of the relationship between response and survival among patients with higher-risk myelodysplastic syndromes treated within azacitidine or conventional care regimens in the randomized AZA-001 trial. *Haematologica*. 2013 Jul;98(7):1067-72.
  29. Itzykson R, Thepot S, Quesnel B, Dreyfus F, Beyne-Rauzy O, Turlure P, Vey N, Recher C, Dartigeas C, Legros L, Delaunay J, Salanoubat C, Visanica S et al. Prognostic factors for response and overall survival in 282 patients with higher-risk myelodysplastic syndromes treated with azacitidine. *Blood*. 2011 Jan 13;117(2):403-11.
  30. Hwang KL, Song MK, Shin HJ, Na HJ, Shin DH, Kim JK, Moon JH, Ahn JS, Song IC, Hong J, Lee GW, Chung JS. Monosomal and complex karyotypes as prognostic parameters in patients with International Prognostic Scoring System higher risk myelodysplastic syndrome treated with azacitidine. *Blood Res*. 2014 Dec;49(4):234-40.

31. Bejar R, Lord A, Stevenson K, Bar-Natan M, Perez-Ladaga A, Zaneveld J, Wang H, Caughey B, Stojanov P, Getz G, Garcia-Manero G, Kantarjian H, Chen R et al. TET2 mutations predict response to hypomethylating agents in myelodysplastic syndrome patients. *Blood*. 2014 Oct 23;124(17):2705-12.
32. Traina F, Visconte V, Elson P, Tabarroki A, Jankowska AM, Hasrouni E, Sugimoto Y, Szpurka H, Makishima H, O'Keefe CL, Sekeres MA, Advani AS, Kalaycio M et al. Impact of molecular mutations on treatment response to DNMT inhibitors in myelodysplasia and related neoplasms. *Leukemia*. 2014 Jan;28(1):78-87.
33. Itzykson R, Kosmider O, Cluzeau T, Mansat-De Mas V, Dreyfus F, Beyne-Rauzy O, Quesnel B, Vey N, Gelsi-Boyer V, Raynaud S, Preudhomme C, Ades L, Fenaux P et al. Impact of TET2 mutations on response rate to azacitidine in myelodysplastic syndromes and low blast count acute myeloid leukemias. *Leukemia*. 2011 Jul;25(7):1147-52.
34. Tobiasson M, McLornan DP, Karimi M, Dimitriou M, Jansson M, Azenkoud AB, Jadersten M, Lindberg G, Abdulkadir H, Kulasekararaj A, Ungerstedt J, Lennartsson A, Ekwall K et al. Mutations in histone modulators are associated with prolonged survival during azacitidine therapy. *Oncotarget*. 2016 Mar 3.
35. Jin J, Hu C, Yu M, Chen F, Ye L, Yin X, Zhuang Z, Tong H. Prognostic value of isocitrate dehydrogenase mutations in myelodysplastic syndromes: a retrospective cohort study and meta-analysis. *PLOS ONE*. 2014;9(6):e100206.
36. Welch JS, Petti AA, Miller CA, Fronick CC, O'Laughlin M, Fulton RS, Wilson RK, Baty JD, Duncavage EJ, Tandon B, Lee YS, Wartman LD, Uy GL et al. TP53 and Decitabine in Acute Myeloid Leukemia and Myelodysplastic Syndromes. *N Engl J Med*. 2016 Nov 24;375(21):2023-36.
37. Jung SH, Kim YJ, Yim SH, Kim HJ, Kwon YR, Hur EH, Goo BK, Choi YS, Lee SH, Chung YJ, Lee JH. Somatic mutations predict outcomes of hypomethylating therapy in patients with myelodysplastic syndrome. *Oncotarget*. 2016 Aug 23;7(34):55264-75.
38. Follo MY, Finelli C, Mongiorgi S, Clissa C, Bosi C, Testoni N, Chiarini F, Ramazzotti G, Baccarani M, Martelli AM, Manzoli L, Martinelli G, Cocco L. Reduction of phosphoinositide-phospholipase C beta1 methylation predicts the responsiveness to azacitidine in high-risk MDS. *Proc Natl Acad Sci U S A*. 2009 Sep 29;106(39):16811-6.
39. Yan P, Frankhouser D, Murphy M, Tam HH, Rodriguez B, Curfman J, Trimarchi M, Geyer S, Wu YZ, Whitman SP, Metzeler K, Walker A, Klisovic R et al. Genome-wide methylation profiling in decitabine-treated patients with acute myeloid leukemia. *Blood*. 2012 Sep 20;120(12):2466-74.
40. Shen L, Kantarjian H, Guo Y, Lin E, Shan J, Huang X, Berry D, Ahmed S, Zhu W, Pierce S, Kondo Y, Oki Y, Jelinek J et al. DNA Methylation Predicts Survival and Response to Therapy in Patients With Myelodysplastic Syndromes. 2010 2010-02-01.
41. Daskalakis M, Nguyen TT, Nguyen C, Guldberg P, Kohler G, Wijermans P, Jones PA, Lubbert M. Demethylation of a hypermethylated P15/INK4B gene in patients with myelodysplastic syndrome by 5-Aza-2'-deoxycytidine (decitabine) treatment. *Blood*. 2002 Oct 15;100(8):2957-64.
42. Gore SD, Baylin S, Sugar E, Carraway H, Miller CB, Carducci M, Grever M, Galm O, Dausies T, Karp JE, Rudek MA, Zhao M, Smith BD et al. Combined DNA methyltransferase and histone deacetylase inhibition in the treatment of myeloid neoplasms. *Cancer Res*. 2006 Jun 15;66(12):6361-9.
43. Raj K, John A, Ho A, Chronis C, Khan S, Samuel J, Pomplun S, Thomas NS, Mufti GJ. CDKN2B methylation status and isolated chromosome 7 abnormalities predict responses to treatment with 5-azacytidine. *Leukemia*. 2007 Sep;21(9):1937-44.
44. Voso MT, Fabiani E, Piciocchi A, Matteucci C, Brandimarte L, Finelli C, Pogliani E, Angelucci E, Fioritoni G, Musto P, Greco M, Criscuolo M, Fianchi L et al. Role of BCL2L10 methylation and TET2 mutations in higher risk myelodysplastic syndromes treated with 5-azacytidine. *Leukemia*. 2011 Dec;25(12):1910-3.

45. Fandy TE, Herman JG, Kerns P, Jiemjit A, Sugar EA, Choi SH, Yang AS, Aucott T, Dausies T, Odchimar-Reissig R, Licht J, McConnell MJ, Nasrallah C et al. Early epigenetic changes and DNA damage do not predict clinical response in an overlapping schedule of 5-azacytidine and entinostat in patients with myeloid malignancies. *Blood*. 2009 Sep 24;114(13):2764-73.
46. Unnikrishnan A, Papaemmanuil E, Beck D, Deshpande NP, Verma A, Kumari A, Woll PS, Richards LA, Knezevic K, Chandrakanthan V, Thoms JAI, Tursky ML, Huang Y et al. Integrative Genomics Identifies the Molecular Basis of Resistance to Azacitidine Therapy in Myelodysplastic Syndromes. *Cell Rep*. 2017 Jul 18;20(3):572-85.
47. Khan R, Schmidt-Mende J, Karimi M, Gogvadze V, Hassan M, Ekstrom TJ, Zhivotovsky B, Hellstrom-Lindberg E. Hypomethylation and apoptosis in 5-azacytidine-treated myeloid cells. *Exp Hematol*. 2008 Feb;36(2):149-57.
48. Chandrakanthan V, Yeola A, Kwan JC, Oliver RA, Qiao Q, Kang YC, Zarzour P, Beck D, Boelen L, Unnikrishnan A, Villanueva JE, Nunez AC, Knezevic K et al. PDGF-AB and 5-Azacytidine induce conversion of somatic cells into tissue-regenerative multipotent stem cells. *Proc Natl Acad Sci U S A*. 2016 Apr 19;113(16):E2306-15.
49. Aimiwu J, Wang H, Chen P, Xie Z, Wang J, Liu S, Klisovic R, Mims A, Blum W, Marcucci G, Chan KK. RNA-dependent inhibition of ribonucleotide reductase is a major pathway for 5-azacytidine activity in acute myeloid leukemia. *Blood*. 2012 May 31;119(22):5229-38.
50. Chiappinelli KB, Strissel PL, Desrichard A, Li H, Henke C, Akman B, Hein A, Rote NS, Cope LM, Snyder A, Makarov V, Buhu S, Slamon DJ et al. Inhibiting DNA Methylation Causes an Interferon Response in Cancer via dsRNA Including Endogenous Retroviruses. *Cell*. 2015 Aug 27;162(5):974-86.
51. Roulois D, Loo Yau H, Singhanian R, Wang Y, Danesh A, Shen SY, Han H, Liang G, Jones PA, Pugh TJ, O'Brien C, De Carvalho DD. DNA-Demethylating Agents Target Colorectal Cancer Cells by Inducing Viral Mimicry by Endogenous Transcripts. *Cell*. 2015 Aug 27;162(5):961-73.
52. Tobiasson M, Abdulkadir H, Lennartsson A, Katayama S, Marabita F, De Paepe A, Karimi M, Krjutskov K, Einarsdottir E, Grovdal M, Jansson M, Ben Azenkoud A, Corddedu L et al. Comprehensive mapping of the effects of azacitidine on DNA methylation, repressive/permissive histone marks and gene expression in primary cells from patients with MDS and MDS-related disease. *Oncotarget*. 2017 Apr 25;8(17):28812-25.
53. Stein EM, DiNardo CD, Pollyea DA, Fathi AT, Roboz GJ, Altman JK, Stone RM, DeAngelo DJ, Levine RL, Flinn IW, Kantarjian HM, Collins R, Patel MR et al. Enasidenib in mutant IDH2 relapsed or refractory acute myeloid leukemia. *Blood*. 2017 Aug 10;130(6):722-31.
54. Martino R, Iacobelli S, Brand R, Jansen T, van Biezen A, Finke J, Bacigalupo A, Beelen D, Reiffers J, Devergie A, Alessandrino E, Mufti GJ, Barge R et al. Retrospective comparison of reduced-intensity conditioning and conventional high-dose conditioning for allogeneic hematopoietic stem cell transplantation using HLA-identical sibling donors in myelodysplastic syndromes. *Blood*. 2006 Aug 1;108(3):836-46.
55. Koenecke C, Gohring G, de Wreede LC, van Biezen A, Scheid C, Volin L, Maertens J, Finke J, Schaap N, Robin M, Passweg J, Cornelissen J, Beelen D et al. Impact of the revised International Prognostic Scoring System, cytogenetics and monosomal karyotype on outcome after allogeneic stem cell transplantation for myelodysplastic syndromes and secondary acute myeloid leukemia evolving from myelodysplastic syndromes: a retrospective multicenter study of the European Society of Blood and Marrow Transplantation. *Haematologica*. 2015 Mar;100(3):400-8.
56. Deeg HJ, Scott BL, Fang M, Shulman HM, Gyurkocza B, Myerson D, Pagel JM, Platzbecker U, Ramakrishnan A, Radich JP, Sandmaier BM, Sorrow M, Stirewalt DL et al. Five-group cytogenetic risk classification, monosomal karyotype, and outcome after hematopoietic cell transplantation for MDS or acute leukemia evolving from MDS. *Blood*. 2012 Aug 16;120(7):1398-408.

57. Cutler CS, Lee SJ, Greenberg P, Deeg HJ, Perez WS, Anasetti C, Bolwell BJ, Cairo MS, Gale RP, Klein JP, Lazarus HM, Liesveld JL, McCarthy PL et al. A decision analysis of allogeneic bone marrow transplantation for the myelodysplastic syndromes: delayed transplantation for low-risk myelodysplasia is associated with improved outcome. *Blood*. 2004 Jul 15;104(2):579-85.
58. Della Porta MG, Alessandrino EP, Bacigalupo A, van Lint MT, Malcovati L, Pascutto C, Falda M, Bernardi M, Onida F, Guidi S, Iori AP, Cerretti R, Marengo P et al. Predictive factors for the outcome of allogeneic transplantation in patients with MDS stratified according to the revised IPSS-R. *Blood*. 2014 Apr 10;123(15):2333-42.
59. Alessandrino EP, Della Porta MG, Bacigalupo A, Van Lint MT, Falda M, Onida F, Bernardi M, Iori AP, Rambaldi A, Cerretti R, Marengo P, Pioltelli P, Malcovati L et al. WHO classification and WPSS predict posttransplantation outcome in patients with myelodysplastic syndrome: a study from the Gruppo Italiano Trapianto di Midollo Osseo (GITMO). *Blood*. 2008 Aug 1;112(3):895-902.
60. Bejar R, Stevenson KE, Caughey B, Lindsley RC, Mar BG, Stojanov P, Getz G, Steensma DP, Ritz J, Soiffer R, Antin JH, Alyea E, Armand P et al. Somatic mutations predict poor outcome in patients with myelodysplastic syndrome after hematopoietic stem-cell transplantation. *J Clin Oncol*. 2014 Sep 1;32(25):2691-8.
61. Yoshizato T, Nannya Y, Atsuta Y, Shiozawa Y, Iijima-Yamashita Y, Yoshida K, Shiraishi Y, Suzuki H, Nagata Y, Sato Y, Kakiuchi N, Matsuo K, Onizuka M et al. Genetic abnormalities in myelodysplasia and secondary acute myeloid leukemia: impact on outcome of stem cell transplantation. *Blood*. 2017 Apr 27;129(17):2347-58.
62. Damaj G, Mohty M, Robin M, Michallet M, Chevallier P, Beguin Y, Nguyen S, Bories P, Blaise D, Maillard N, Rubio MT, Fegueux N, Cornillon J et al. Upfront allogeneic stem cell transplantation after reduced-intensity/nonmyeloablative conditioning for patients with myelodysplastic syndrome: a study by the Societe Francaise de Greffe de Moelle et de Therapie Cellulaire. *Biol Blood Marrow Transplant*. 2014 Sep;20(9):1349-55.
63. Oran B, Kongtim P, Popat U, de Lima M, Jabbour E, Lu X, Chen J, Rondon G, Kebriaei P, Ahmed S, Andersson B, Alousi A, Ciurea S et al. Cytogenetics, donor type, and use of hypomethylating agents in myelodysplastic syndrome with allogeneic stem cell transplantation. *Biol Blood Marrow Transplant*. 2014 Oct;20(10):1618-25.
64. Gerds AT, Gooley TA, Estey EH, Appelbaum FR, Deeg HJ, Scott BL. Pretransplantation therapy with azacitidine vs induction chemotherapy and posttransplantation outcome in patients with MDS. *Biol Blood Marrow Transplant*. 2012 Aug;18(8):1211-8.
65. Damaj G, Duhamel A, Robin M, Beguin Y, Michallet M, Mohty M, Vigouroux S, Bories P, Garnier A, El Cheikh J, Bulabois CE, Huynh A, Bay JO et al. Impact of azacitidine before allogeneic stem-cell transplantation for myelodysplastic syndromes: a study by the Societe Francaise de Greffe de Moelle et de Therapie-Cellulaire and the Groupe-Francophone des Myelodysplasies. *J Clin Oncol*. 2012 Dec 20;30(36):4533-40.
66. Schroeder T, Rachlis E, Bug G, Stelljes M, Klein S, Steckel NK, Wolf D, Ringhoffer M, Czibere A, Nachtkamp K, Dienst A, Kondakci M, Stadler M et al. Treatment of acute myeloid leukemia or myelodysplastic syndrome relapse after allogeneic stem cell transplantation with azacitidine and donor lymphocyte infusions--a retrospective multicenter analysis from the German Cooperative Transplant Study Group. *Biol Blood Marrow Transplant*. 2015 Apr;21(4):653-60.
67. Tobiasson M, Olsson R, Hellstrom-Lindberg E, Mattsson J. Early detection of relapse in patients with myelodysplastic syndrome after allo-SCT. *Bone Marrow Transplant*. 2011 May;46(5):719-26.
68. Platzbecker U, Wermke M, Radke J, Oelschlaegel U, Seltmann F, Kiani A, Klut IM, Knoth H, Rollig C, Schetelig J, Mohr B, Graehlert X, Ehninger G et al. Azacitidine for treatment of imminent relapse in MDS or AML patients after allogeneic HSCT: results of the RELAZA trial. *Leukemia*. 2012 Mar;26(3):381-9.